

STRUCTURES OF SOME MINOR PTEROCARPANS OF *NEORAUTANENIA EDULIS*

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Abstract—The constitution of five new compounds, (–)-neorautenol, (–)-neodunol, (–)-homoeudiol, neorauten and neodulen, has been established in the root bark of *Neorautanenia edulis*

INTRODUCTION

THE POSITION of the genus *Neorautanenia* in the family Leguminosae suggests that these plants could be sources of isoflavonoids and rotenoids and this apparently correlates with claims that some *Neorautanenia* species have been used extensively throughout Central and Southern Africa as fish poisons.¹ *N. edulis* in particular has been the subject of several investigations^{2–5} concerned with the isolation of the active principles responsible for these properties. The present communication discusses the isolation, structural elucidation and biogenetic significance of the concurrence of pterocarpanoids and 6a,11a-dehydropterocarpanes in this plant.

RESULTS AND DISCUSSION

Neorautenol (1)

The compound, $[\alpha]_D^{22} - 273^\circ$ ($c = 0.3$, CHCl_3), was shown by analysis, and MS to have a formula $\text{C}_{20}\text{H}_{18}\text{O}_4$. The presence of a single OH group was shown by the formation of a mono-methyl ether and mono-acetate. NMR comparison (Table 1) of neorautenol with other pterocarpanes indicates that these compounds have the same A/B/C/D ring system and that structural differences concern the accompanying substitution pattern. The protons in the *para*-position on ring A resonate as singlets and the three ring-D aromatic protons are readily analysed in terms of an ABX system. The characteristic ABCD pattern of pterocarpanes⁶ (arising from the 6a-, 11a-, 6_{eq}- and 6_{ax}-protons) is easily discerned, although the signal from the 6_{ax}-proton is partly obscured by the methoxy-resonances.

The MS of the resulting methoxylated isoflavan (7) agrees well with the established pattern for isoflavans^{7,8} on the assumption that ring A has a γ,γ -dimethylchromon group and

¹ WATT, J. M. and BREYER-BRANDWIK, M. G. (1962) *Medical and Poisonous Plants of Southern and Eastern Africa*, 2nd Edit., pp 636, E and S Livingstone, London

² RALL, G. J. H., ENGELBRECHT, J. P. and BRINK, A. I. (1972). *J. S. African. Chem. Inst.* **25**, 131.

³ VAN DUUREN, B. L. (1961) *J. Org. Chem.* **26**, 5013

⁴ VAN DUUREN, B. L. and GROENEWOLD, P. W. G. (1950). *J. S. African. Chem. Inst.* **3**, 29

⁵ VAN DUUREN, B. L. and GROENEWOLD, P. W. G. (1950). *J. S. African. Chem. Inst.* **3**, 35

⁶ PACHLER, K. G. R. and UNDERWOOD, W. G. E. (1967) *Tetrahedron* **23**, 1817

⁷ PELTER, A., STANTON, P. and BARBER, M. (1965) *J. Heterocyclic Chem.* **2**, 262

⁸ PELTER, A. and AMENECHE, P. I. (1969) *J. Chem. Soc. (C)* 887

TABLE 1. ASSIGNMENTS OF CHEMICAL SHIFTS (τ IN PPM) IN THE NMR SPECTRA OF SOME PTROCARPANS AND RELATED COMPOUNDS

	H6a*	H6 _{ac} *	H6 _q *	H11a†	H2	H3	H4	H1	H4	H6	H7‡	H8§	H9	H10
Neorautenol 1	6.48	6.38	5.78	4.51	8.57 \times 3H 8.59 \times 3H	4.44 <i>d</i> <i>I</i> 10.0	3.67 <i>d</i> <i>I</i> 10.0	2.84 \times	3.58 \times		2.91 <i>I</i> 9.0	3.64 <i>I</i> 9.0 <i>I</i> 2.5		3.60 <i>I</i> 2.5
1a	6.55	6.42	5.85	4.59	8.60 <i>br</i> \times 6H	4.44 <i>d</i> <i>I</i> 10.0	3.68 <i>d</i> <i>I</i> 10.0	2.89 \times	3.58 \times		2.92 <i>I</i> 9.0	3.58 <i>I</i> 9.0 <i>I</i> 2.5	6.30 \times OCH ₃	3.58 <i>I</i> 2.5
1b	6.50	6.33	5.77	4.53	8.58 <i>br</i> \times 6H	4.46 <i>d</i> <i>I</i> 10.0	3.68 <i>d</i> <i>I</i> 10.0	2.88 \times	3.60 \times		2.80 <i>I</i> 9.0	3.38 <i>I</i> 9.0 <i>I</i> 2.5	7.74 \times OCOCH ₃	3.39 <i>I</i> 2.5
Neodunol 2	6.37	6.27	5.68	4.30	2.42 <i>d</i> <i>I</i> 2.2	3.25 <i>d</i> <i>I</i> 2.2		2.23 \times	2.87 \times		2.89 <i>I</i> 8.5	3.61 <i>I</i> 8.5 <i>I</i> 2.5		3.60 <i>I</i> 2.5
2a	6.37	6.18	5.70	4.30	2.44 <i>d</i> <i>I</i> 2.2	3.26 <i>d</i> <i>I</i> 2.2		2.27 \times	2.88 \times		2.83 <i>I</i> 9.0	3.53 <i>I</i> 9.0 <i>I</i> 2.5	6.23 \times OCH ₃	3.52 <i>I</i> 2.5
2b	6.35	6.25	5.68	4.29	2.44 <i>d</i> <i>I</i> 2.2	3.28 <i>d</i> <i>I</i> 2.2		2.27 \times	2.89 \times		2.77 <i>I</i> 9.0	3.36 <i>I</i> 9.0 <i>I</i> 2.5	7.75 \times OCOCH ₃	3.38 <i>I</i> 2.5
Homoedudiol 3	6.48	6.48	6.33	5.52	4.51	8.22 <i>br</i> \times 6H ^{9,10}	6.73 <i>d</i> <i>I</i> 7.0 2H	4.74 <i>t</i> <i>I</i> 7.0 1H	3.59 \times		2.59 <i>I</i> 8.5	3.43 <i>I</i> 8.5 <i>I</i> 2.5		3.55 <i>I</i> 2.5
3a	6.52	6.38	5.74	4.53	8.27 <i>br</i> \times 6H	6.73 <i>d</i> <i>I</i> 7.0 2H	4.74 <i>t</i> <i>I</i> 7.0 1H	2.73 \times	3.56 \times		2.68 <i>I</i> 8.5	3.56 <i>I</i> 8.5 <i>I</i> 2.5	6.26 \times 3H 6.22 \times 3H	3.53 <i>I</i> 2.5
Neorautenol Methylether 4a					2.48 <i>d</i> <i>I</i> 2.2	3.29 <i>d</i> <i>I</i> 2.2		2.36 \times	2.93 \times	4.45 \times 2H	2.76 <i>I</i> 8.5	3.14 <i>I</i> 8.5 <i>I</i> 2.5	6.13 \times OCH ₃	2.92 <i>I</i> 2.5
4b					2.49 <i>d</i> <i>I</i> 2.2	3.31 <i>d</i> <i>I</i> 2.2		2.35 \times	2.95 \times	4.47 \times 2H	2.68 <i>I</i> 8.5	3.03 <i>I</i> 8.5 <i>I</i> 2.5	7.73 \times OCOCH ₃	2.71 <i>I</i> 2.5
Neodulcin 5					2.49 <i>d</i> <i>I</i> 2.2	3.31 <i>d</i> <i>I</i> 2.2		2.37 \times	2.94 \times	4.49 \times 2H			OCH ₃ O 4.02 \times	3.25 \times

* A multiplet centred at the τ value given† A doublet at the τ value given, showing signs of further splitting

‡ The A proton in an ABX system appearing as a doublet

§ The B proton in an ABX system appearing as a quartet centred at the τ value given

| The X proton in an ABX system appearing as a doublet

ring B two methoxy substituents [VII $\xrightarrow{\text{RDA}}$ m/e 190 (14), m/e 164 (100)] This evidence suggests that neorautenol is the phenolic pterocarpin (1). This was confirmed by the conversion of 1 to homoedudiol (3).

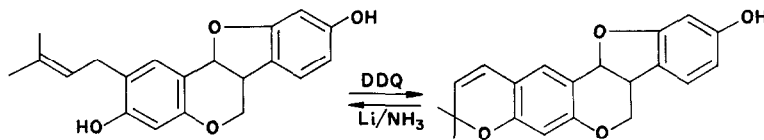
Homoedudiol (3)

This compound, C₂₀H₂₀O₄, had ν_{max} (CHCl₃) 3370 cm⁻¹, M⁺ 324. The presence of two phenolic OH groups as indicated by the formation of a dimethyl ether. M⁺ 352. Reduction of neorautenol (1) with lithium and ammonia afforded a major product identical to the substance (3). This was confirmed by the conversion of homoedudiol (3) to (1) with DDQ.

⁹ OLLIS, W. D., RAMSAY, M. V. J., SUTHERLAND, I. O. and MONGKOLSUK, S. (1965) *Tetrahedron* **21**, 1453.

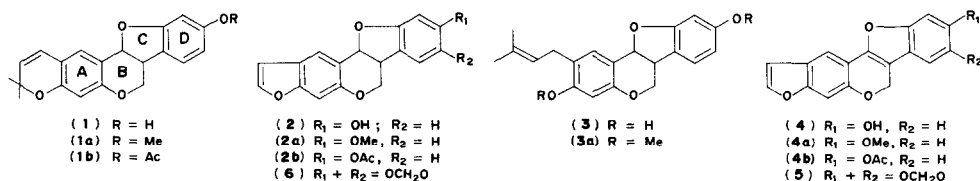
¹⁰ BATES, R. B. and GALT, D. M. (1960) *J. Am. Chem. Soc.* **82**, 5749.

(Scheme 1). Structure (3) was hence indicated, the *ortho*-(3,3-dimethylallyl)-phenol explaining the facile loss of isobutene ($M^+ - 55$) in the MS.¹⁷



Neodunol (2)

The third compound, formulated as the new pterocarpan (2), was isolated as the acetate and the free phenol was regenerated by hydrolysis with ammonium hydroxide. The presence of a benzofuran ring system¹¹ was indicated by characteristic UV and NMR spectra [λ_{\max} 246 nm (log ϵ 4.34); τ 2.42, *d*, *J* 2.2 Hz, H2' and τ 3.25, *d*, *J* 2.2 Hz, H3'], while the aromatic region in the NMR spectrum (Table 1) of neodunol suggests a similar oxygenation pattern as in neorauteenol (1) and homoeudiol (3). The new compound is hence identified as the phenolic pterocarpan (2) and its MS fully supports this structure with prominent ions at *m/e* 147 ($M^+ - 133$), 263 ($M^+ - 17$, metastable at 246.8) and 171 ($M^+ - 1 - 108$).



The pterocarpenes, neorauteen (4) and neoduleen (5)

The UV spectra of neorauteen (4), C₁₇H₁₀O₄ and neoduleen (5), C₁₈H₁₀O₅, were very similar to that of dehydrohomoptercarpin (8)¹² and flemichapparin-B (9)¹³ (Table 2).

TABLE 2 UV ABSORPTION SPECTRA OF NEODULEEN, NEORAUTEEN AND RELATED PTEROCARPENES

	λ_{\max} nm (log ϵ) in EtOH				
Neoduleen (5)	224 (4.30),	252 (4.16),	294 (3.84),	346 (4.28),	364 (4.18)
Neorauteen (4)	222 (4.35);	250 (4.17),	294 (3.82),	344 (4.36),	360 (4.21)
Dehydrohomoptercarpin (8) ¹²		244 (4.23),	303 (3.81),	342 (4.61)	
Flemichapparin-B (9) ¹³	215 (4.40),	244 (4.20),	291 (3.80);	339 (4.58),	358 (4.16)

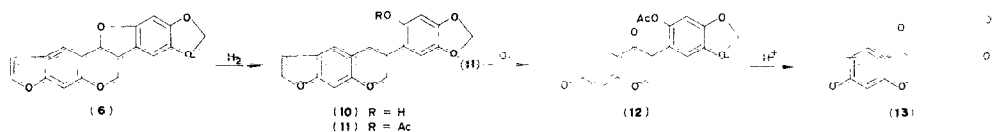
Compound (5) gave a positive Labat test for a methylenedioxy group. The IR showed no OH or carbonyl absorption, but a band at 1660 cm⁻¹ indicated olefinic unsaturation. This was reflected in the UV spectrum which showed high intensity absorption with maxima up to 364 nm (log ϵ 4.18). The NMR spectrum contained a peak at τ 4.02 expected for a methylenedioxy group. The aromatic region of the spectrum consisted of 4 singlets (τ 2.37, H1; 2.94, H4; 3.25, H10; 2.97, H7) and 2 doublets [τ 2.42, 3.36; *J* 2.2 Hz (H2' and H3')] indicative of a furan ring, while the remainder of the spectrum consisted of a singlet representing two protons at τ 4.49 (CH₂ at C-6). From this evidence compound (5) was

¹¹ WESSLEY, F and KOTLAN, J (1955) *Monatsh* **86**, 431

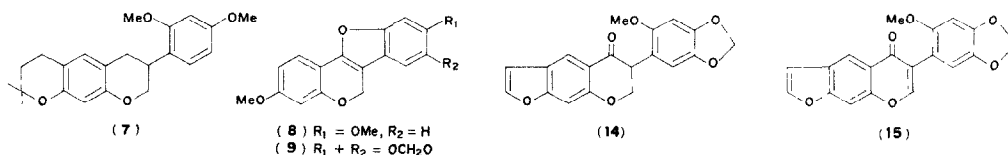
¹² HARPER, S H, KEMP, A D, UNDERWOOD, W G E and CAMPBELL, R V M (1969) *J Chem Soc (C)*, 1109

¹³ CHANDHURY, N A and GUPTA, P K (1970) *Chem Ind.*, 745

considered to be 2,3,4,5-furo-8,9-methylenedioxyneodulol (6) and neodulol (6) was obtained by the direct synthetic cyclization of neodulin (6) and neoduleen (5) (Scheme 2). Hydrogenolysis of (6) gave the isoflavanone (10). Oxidation of the corresponding acetate (11) afforded the isoflavanone (12) which undergoes cyclization to yield neodulol (13). Treatment of (13) with DDQ gave a compound identical with the natural product.



Neorauteen (4) was isolated as the acetate and methyl ether. From the spectrometric information it was evident that a very close relationship exists between neorauteen and neoduleen (5). The NMR spectra of (4a) and (4b) were most informative where an ABX system (τ 2.76, *d*, *J* 8.5 Hz, H7; τ 3.14, *q*, *J*₁ 8.5 Hz, *J*₂ 2.5 Hz, H8; τ 2.92, *d*, *J* 8.5 Hz, H10) in the aromatic region was discernable as the only difference. Neorauteen was thus the phenolic pterocarpen with constitution (4).



The co-occurrence of dehydroneotenone (15),¹⁴ neotenone (14),¹⁴ neodulin (6)¹³ and neoduleen (5) in the same plant provides circumstantial evidence for the hypothesis^{15,16} that pterocarpenes, via direct oxidative cyclization of 2'-hydroxyisoflavanones, are intermediates in the biosynthesis of pterocarpanes. This was also shown by the chemical conversion of the isoflavanone (12) to the pterocarpan (13) (Scheme 2).

EXPERIMENTAL

Mps are uncorrected. IR spectra were recorded in KBr discs and UV spectra in EtOH. NMR were determined using a Varian T-60 spectrometer and MS were recorded on an AEI MS-9 instrument. Chemical shifts are expressed in ppm downfield from TMS. Abbreviations: *s* = singlet, *d* = doublet, *m* = multiplet and *br* = broad. Preparative TLC was carried out on silica gel GF₂₅₄ plates (1 mm thick).

Isolation of compounds. Dried, milled bark (5 kg) was extracted in 120 g portions with refluxing hexane for 24 hr. The combined hexane soln was concentrated to give an extract (47 g) which was carefully chromatographed on silica gel (170–230 mesh) with C₆H₆–CHCl₃ (with increasing concentrations of CHCl₃ from 5 to 60%) as solvent. Combined fractions were submitted to preparative TLC to purify the compounds: (a) Neorauteenol (1) gave colourless needles (70 mg) from MeOH, m.p. 168.5–170° after TLC in hexane–Me₂CO (13:7), *R_f* 0.61, *M*⁺ 322, [α]_D²⁵ – 188.2 (c 1.5, CHCl₃); (b) Neodulin (2), TLC *R_f* 0.54 in hexane–Me₂CO (13:7) was crystallized from C₆H₆–hexane (9:1) to give colourless needles (43 mg), m.p. 170.5–171.5°, *M*⁺ 280, [α]_D²⁵ – 284.9 (c 0.8, CHCl₃); (c) Homoedudiol (3) was obtained as a colourless oil (57 mg) after purification by TLC in CHCl₃–LiOAc (13:7), *R_f* 0.53, *M*⁺ 324, [α]_D²⁵ – 246.1 (c 0.8, CHCl₃); (d) After TLC in CHCl₃–MeOH (96:4), *R_f* 0.53, neorauteen (4) (73 mg) was crystallized from MeOH to give white plates, m.p. 202.5–204.0°, *M*⁺ 278; (e) Neoduleen (5) was obtained by TLC in C₆H₆–hexane (3:1), *R_f* 0.72. Crystallization from MeOH afforded white needles (80 mg), m.p. 221.5–223.0°, *M*⁺ 306.

¹⁴ BRINK, C. V. D. M., DEKKER, I. I., HANEKOM, E. C., MEJERING, D. H. and RALL, G. I. H. (1965), *J. S. African Chem. Inst.* **18**, 21.

¹⁵ ADITYACHANDHURY, N. and GUPTA, P. K. (1970) *Chem. Ind.* 1113.

¹⁶ WENG, E. (1976), In: *Progress in Chemistry of Organic Natural Products* (HERZ, W., GRISEBACH, H. and SCOTT, A. I., eds) pp. 28, 57, Springer, New York.

¹⁷ BALLANTRINE, I. A., FRANCIS, D. I., HASSALL, C. H. and WRIGHT, I. L. C. (1970), *J. Chem. Soc. (C)*, 1175.

Acetylation and methylation methods A soln of the compound (20–40 mg) in dry pyridine (2 ml) and Ac_2O (3 ml) was heated for 1 hr at 100° and the soln was poured into ice (20 g). The ppt. was extracted with Et_2O (3×50 ml), washed with cold M HCl (2×20 ml) and H_2O (3×20 ml). Evaporation of the solvent from the dried (MgSO_4) extract afforded the acetates. Me_2SO_4 (1 ml) was added slowly over a period of 1 hr to a refluxing mixture of the compound (20–40 mg) in dry Me_2CO (50 ml) and anhyd. K_2CO_3 (1.5 g). The mixture was stirred for 1 hr while heating continued. After cooling 5% NaOH (20 ml) was added. The soln was allowed to stand overnight, and the resulting ppt. extracted with Et_2O (3×50 ml). Evaporation of the solvent from the dried (MgSO_4) extract gave the methyl ethers.

Neorautenol monomethylether (1a) Methylation of (1) (30 mg) afforded (1a) (24 mg) as a colourless oil, M^+ 336.

Neorautenol acetate (1b) Acetylation of (1) (25 mg) gave (1b) (20 mg) as colourless needles, m.p. $102.5\text{--}104^\circ$, M^+ 380.

2'',2''-Dimethyl-6,7,5'',6''-chroman-2',4'-dimethoxyisoflavan (7) Hydrogenolysis of neorautenol (1) (50 mg) in EtOAc (50 ml) over 10% Pd catalyst (on carbon support), according to standard procedure gave the corresponding phenol. Due to the instability of this compound it was methylated to give (7) as a colourless oil, M^+ 354, τ (100 MHz) 5.67, q, J_1 10 Hz, J_2 3.5 Hz, 2-H, 6.00, t, J 10 Hz, 2-H, 6.20–6.56, m, 3-H, 7.06, br s, 4-H, 7.23, br s, 4-H, 6.25, s, OMe, 6.26, s, OMe, 3.53, d, J 2.5 Hz, 3'-H, 3.49, q, J_1 9 Hz, J_2 2.5 Hz, 5'-H, 2.91, d, J 9 Hz, 6'-H, 8.58, br s, 2''-C (CH_3)₂, 8.24, t, J 6.5 Hz, 3''-CH₂, 7.24, t, J 6.5 Hz, 4''-CH₂, 3.73, s, 5-H, 3.78, s, 8-H.

Birch reduction of neorautenol (1) Li (50 mg) was added to a soln of (1) (50 mg) in ethyleneglycoldimethylether (20 ml) and liquid NH_3 (30 ml). After 3 min, H_2O (30 ml) was added. The organic phase was evaporated under red. pres. after acidification (6 N HCl). The mixture was extracted with Et_2O (3×50 ml) and evaporation of the solvent gave a mixture of compounds which was separated on TLC in hexane– Me_2CO (13/7) to give (3) (15 mg) as an oil, M^+ 324, identical to an authentic sample.

Homoedudiolmethylether (3a), (3) (30 mg) was treated with an excess of CH_2N_2 in ether at 0° for 24 hrs. Evaporation of the solvent yielded (3a) as a colourless oil, M^+ 352, identical to an authentic specimen.

Reaction of homoedudiol (3) with DDQ A mixture of (3) (30 mg) and DDQ (25 mg) in Na dried C_6H_6 (20 ml) was refluxed for 30 min. The reaction mixture was filtered and chromatographed on a short alumina column. After evaporation of the solvent the residue was submitted to preparative TLC in hexane– Me_2CO (13/7), R_f 0.61, to give neorautenol (1) (12 mg) as colourless needles, m.p. $168.5\text{--}170^\circ$ from MeOH , identical to an authentic sample.

Neodunol methylether (2a) Methylation of (2) (20 mg) gave (2a) (14 mg) as colourless needles (C_6H_6), m.p. $158\text{--}159^\circ$, M^+ 294.

Neodunol acetate (2b) Acetylation of (2) (20 mg) with Ac_2O and dry pyridine according to standard procedure afforded (2b) (16 mg) as plates, m.p. $136\text{--}137^\circ$, M^+ 338. Hydrolysis with 5% NH_4OH of (2b) (25 mg) gave neodunol (2) (16 mg) as colourless needles C_6H_6 –hexane (9/1), m.p. $170.5\text{--}171.5^\circ$, M^+ 280.

2'',3''-Dihydro-6,7,4'',5''-furo-2'-acetoxy-4',5'-methylenedioxyisoflavan (11) Hydrogenolysis of neodulin (6) (900 mg) in EtOAc – HOAc (1/1) (50 ml) with 10% Pd/C (500 mg) at 60° for 24 hrs afforded a mixture of the phenolic isoflavan (10) and starting material. After filtration and evaporation of the solvent under red. pres. the crude mixture was acetylated. After work-up the mixture was separated by TLC in CHCl_3 to give (11) (320 mg) as colourless needles, m.p. $144\text{--}145.5^\circ$, M^+ 354, τ (100 MHz) 7.73, s, OCOCH_3 , 7.23, s, 4-H, 7.12, s, 4-H, 6.93, t, J 8.2 Hz, 3'-H, 6.08, t, J 10 Hz, 2-H, 5.77, q, J_1 10 Hz, J_2 3.5 Hz, 2-H; 5.48, t, J 8.2 Hz, 2''-H, 4.05, s, OCH_2O , 3.65, s, 3'-H, 3.40, s, 8-H, 3.33, s, 6'-H, 2.75, s, 5-H.

2'',3''-Dihydro-6,7,4'',5''-furo-2'-acetoxy-4',5'-methylenedioxyisoflavanone (12) A soln of the isoflavan (11) (350 mg) in Me_2CO (50 ml) was oxidized by the dropwise addition of 5% aq. KMnO_4 (40 ml) over a period of 4 hr. H_2O (50 ml) was added before SO_2 was passed through the soln. The H_2O layer was extracted with Et_2O (4×50 ml) after evaporation of the Me_2CO . The residue from the Et_2O extract was separated by TLC in CHCl_3 to give (12) (100 mg) as colourless needles (MeOH), m.p. $180.5\text{--}182^\circ$, M^+ 368, τ (60 MHz) 7.78, s, OCOCH_3 , 6.87, t, J 8.2 Hz, 3'-H, 5.98, d, J 8 Hz, 3-H, 5.56, d, J 8 Hz, 2-H, 5.35, d, J 8.2 Hz, 2''-H, 4.05, s, OCH_2O , 3.65, s, 3'-H, 3.33, s, 6'-H, 3.33, s, 8-H, 2.22, s, 5-H.

2',3'-Dihydroneodulen (13) A soln of (12) (100 mg) in HOAc (8 ml) and conc. HCl (4 ml) was heated for 30 min at 100° . The resulting white ppt. was filtered and crystallized from MeOH to give colourless needles, m.p. $194\text{--}195.5^\circ$, of (13) (67 mg).

Neodulen (5) (13) (60 mg) and DDQ (40 mg) in dry C_6H_6 (30 ml) was stirred under reflux for 2 hrs. After filtration the mixture was separated by TLC in C_6H_6 –hexane (3/2), R_f 0.72, to give (5) (15 mg) as colourless needles, m.p. $222\text{--}223^\circ$, M^+ 306, identical with an authentic sample.

Neorauteen methylether (4a) Methylation of (4) (20 mg) afforded (4a) (13 mg) as a white powder, m.p. $200\text{--}201.5^\circ$, M^+ 292.

Neorauteen acetate (4b) Acetylation of (4) (20 mg) gave (4b) (16 mg) as colourless needles, m.p. $197.5\text{--}199^\circ$, M^+ 336.

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